

THE REVISED STRUCTURE OF  
BOTTROMYCIN A2

Sir:

During the studies on application of chemical ionization (CI) mass spectrometry to antibiotic research, we found that the fragmentation pattern of the spectrum of bottromycin A2 cannot be explained by its proposed structure<sup>1)</sup> (Fig. 1). This prompted us to reinvestigate its chemical structure. In this communication, the revised structure of bottromycin A2 deduced mainly by mass spectrometric analyses, is presented.

The CI mass spectrum of bottromycin A2 together with the electron impact (EI) mass spectrum are shown in Fig. 2. There exist only five significant peaks over  $m/e$  100 in the CI mass spectrum [ $m/e$  823 ( $M + 1$ ), 654, 476, 348 and 170]. These peaks can be assigned to the fragment ions of the proposed structure shown in Fig. 1. For a linear structure of bottromycin A2, one would anticipate additional fragment ions, due to the cleavage of other peptide bonds. Thus, the CI mass spectrum suggests that the fragment ion

$m/e$  476 is cyclic. The molecular formula of bottromycin A2 can be assigned to  $C_{42}H_{62}N_9O_7S$  by high resolution EI mass spectrometry, observed:  $m/e$  822.4411, error  $-4.7$  millimass units (Hitachi RMU-7M spectrometer with 002 data processing system), which is in accord with earlier report<sup>1)</sup>.

The  $^1H$ -NMR spectrum of bottromycin A2 indicated that there is no olefinic proton. The resonance at  $\delta$  6.9 (doublet,  $J = 9$  Hz) in  $CDCl_3$ , which was originally assigned to the olefinic proton of  $\Delta^1$ -isocaproic acid moiety in the proposed structure, can be assigned to an NH proton, because it rapidly disappeared by addition of  $D_2O$ . The  $^{13}C$ -NMR spectrum (Fig. 3) also confirmed the absence of such olefinic carbons. The  $^1H$ - and  $^{13}C$ -NMR spectra rather suggested the presence of two tertiary butyl groups. There is an 18 H-singlet peak at  $\delta$  1.00 in the  $^1H$ -NMR spectrum and 2 quaternary carbon resonances at  $\delta$  33.1 and 35.6, and a very strong methyl carbon resonance at  $\delta$  27.9, of which intensity is about 6 times stronger than a nearby methyl carbon resonance, in the  $^{13}C$ -NMR spectrum.

Fig. 1. The previous structure of bottromycin A2

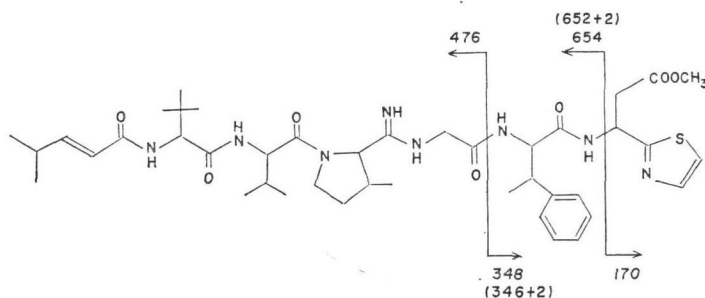


Fig. 2. The CI and EI mass spectra of bottromycin A2 (Hitachi RMU-6M spectrometer)

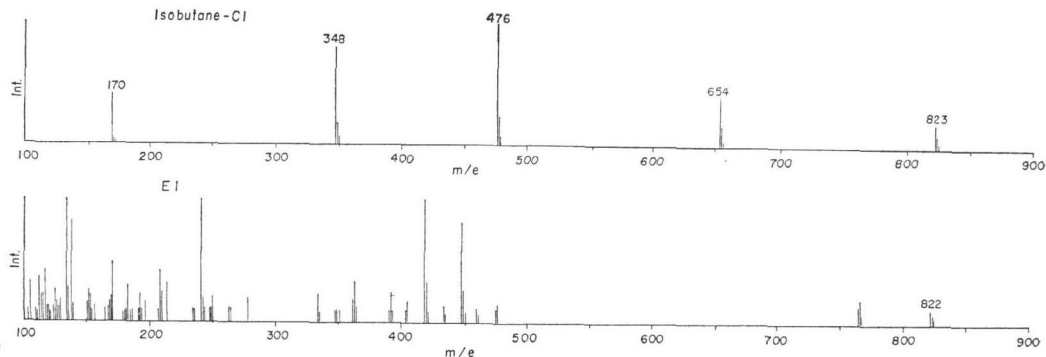
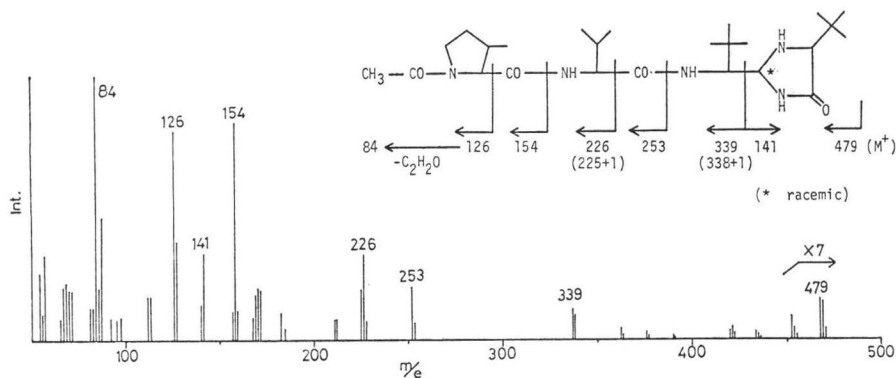


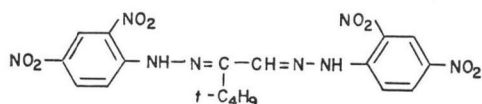


Fig. 4. The EI mass spectrum of the tetrahydrotetrapeptide



ment of the fragment ions was verified by their elemental compositions obtained by high resolution mass spectrometry (Table 1). The rest of the molecule should be an imidazolidone containing another *t*-butyl group ( $m/e$  141, see Fig. 4 and Table 1), which is derived by reduction of the imidazolone in the tetrapeptide.

The imidazolidone is a potential aldehyde. Thus, the tetrahydroderivative was hydrolyzed in 6N HCl in the presence of 2,4-dinitrophenylhydrazine to catch the yielded aldehyde. The hydrolyzate contained an orange-colored substance together with DMAB, valine and 3-methylproline. The molecular formula,  $C_{18}H_{18}N_8O_8$ , of the colored substance was established by high resolution mass spectrometry, observed:  $m/e$  474.1274, error + 2.8 millimass units. The  $^1H$ -NMR spectrum in  $CDCl_3$  showed the presence of a *t*-butyl [ $\delta$  1.38 (9H, singlet)], a methine [ $\delta$  7.94 (1H, singlet)], two 2,4-dinitrophenyl (6 protons of a pair of 1,2,4-substituted benzene at  $\delta$  8.20, 8.28, 8.38, 8.50, 9.13 and 9.15) and two hydrogen-bonded NH groups [ $\delta$  11.41 (1H, singlet), 13.31 (1H, singlet)]. Thus, the structure of the colored substance was determined to be the osazone of *t*-butylglyoxal.



If the tetrahydroderivative has a latent 2-amino-3,3-dimethylbutyraldehyde moiety, it will give 3,3-dimethyl-2-oxo-butyl alcohol by prototropy and deaminative hydration during acid

Table 1. The elemental compositions of the fragment ions shown in Fig. 4

$m/e$ (observed)	Elemental composition	Error (millimass unit)
84.0809	$C_5H_{10}N_1$	-0.2
126.0901	$C_7H_{12}N_1O_1$	-1.6
141.1028	$C_7H_{13}N_2O_1$	+0.1
154.0864	$C_8H_{12}N_1O_2$	-0.2
255.1552	$C_{12}H_{21}N_2O_2$	-4.9
226.1687	$C_{12}H_{22}N_2O_2$	+0.7
253.1547	$C_{13}H_{21}N_2O_3$	-0.3
339.2474	$C_{18}H_{33}N_3O_3$	-4.5
479.3451	$C_{25}H_{45}N_5O_4$	-1.7

hydrolysis. The osazone of *t*-butylglyoxal is derived from the latter in the presence of the phenylhydrazine. The similar reaction is reported by TATSUTA *et al.*<sup>6)</sup>, that is: 2-oxo-3-phenylpropyl alcohol is yielded from the phenylalaninal moiety of chymostatin by acid hydrolysis. Thus, the presence of the imidazolidone was confirmed by isolation of the osazone, and the structure of the tetrahydroderivative was established as shown in Fig. 4.

The  $^1H$ -NMR spectrum of the diastereoisomeric mixture of the tetrahydroderivative showed that the N-acetyl group appeared separately at  $\delta$  1.83 and 1.92 with almost equal intensity, though the acetyl group is far from the racemic carbon. It suggests the presence of intramolecular hydrogen-bondings to keep these groups in proximity. Formation of the fragment ions  $m/e$  226 (225+1) and 339 (338+1) also could be explained by a strong intramolecular hydrogen-bonding between the carbonyl group of 3-

methylprolyl moiety and an NH proton of the imidazolidone with ten-atoms ring.

The structures of the tetrapeptide and bottromycin A2 are presented as shown in Fig. 5\*. It must be noticed that the source of DMAB derived by acid hydrolysis of the tetrapeptide is different from that of its tetrahydroderivative. The CI mass spectrum of bottromycin B2<sup>7)</sup> gave five significant peaks at *m/e* 809 (M+1), 640, 462, 348 and 170. Therefore, the structure of B2 is assigned to be the structure shown in Fig. 5, in which the 3-methylprolyl moiety is substituted by prolyl moiety<sup>7)</sup>. For the structures of the other components<sup>7,8)</sup> of bottromycins, the reinvestigation should be necessary. But the samples are not available now.

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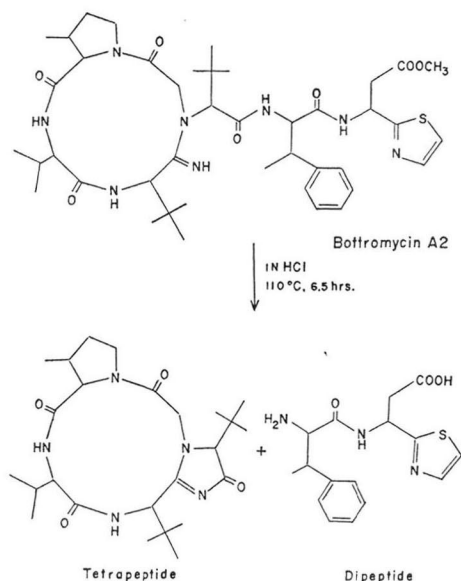
#### References

- 1) NAKAMURA, S. & H. UMEZAWA: The structure of bottromycin A2, a new component of bottromycins. *Chem. Pharm. Bull.* 14: 981~986, 1966
- 2) WAISVISZ, J. M.; M. G. VAN DER HOEVEN & B. TE NIJENHUIS: The structure of the sulfur-containing moiety of bottromycin. *J. Amer. Chem. Soc.* 79: 4524~4527, 1957

\*  $\Delta^1$ -Isocaproic acid unit in bottromycin A2 and the tetrapeptide in the previous structures was deduced from the isolation of isobutylaldehyde by ozonolysis. This isobutylaldehyde might be introduced from the previously used apparatus. The reexamination did not give any volatile carbonyl compound by the ozonolysis of bottromycin A2 and the tetrapeptide.

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Fig. 5. The revised structure of bottromycin A2 and its mild acid hydrolysis products.



- 3) NAKAMURA, S.; T. CHIKAIKE, H. YONEHARA & H. UMEZAWA: Isolation, characterization and structural elucidation of new amino acids from bottromycin A. *Chem. Pharm. Bull.* 13: 599~602, 1965
- 4) WAISVISZ, J. M.; M. G. VAN DER HOEVEN, J. F. HÖLSCHER & B. TE NIJENHUIS: Bottromycin. II. Preliminary degradation studies. *J. Amer. Chem. Soc.* 79: 4522~4524, 1957
- 5) WILCHEK, M.; S. SARID & A. PATCHORNIK: Use of sodium in liquid ammonia for cleavage of N-proline peptides. *Biochim. Biophys. Acta* 104: 616~618, 1965
- 6) TATSUTA, K.; N. MIKAMI, K. FUJIMOTO, S. UMEZAWA, H. UMEZAWA & T. AOYAGI: The structure of chymostatin, a chymotrypsin inhibitor. *J. Antibiotics* 26: 625~646, 1973
- 7) NAKAMURA, S.; T. YAJIMA, YOUNG-CHI LIN & H. UMEZAWA: Isolation and characterization of bottromycins A2, B2, C2. *J. Antibiotics, Ser. A* 20: 1~5, 1967
- 8) NAKAMURA, S.; N. TANAKA & H. UMEZAWA: Bottromycin A1, A2 and their structures. *J. Antibiotics, Ser. A* 19: 10~12, 1966